

Receptor binding specificity and sequence comparison of a novel avian-origin H7N9 virus in China

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ABSTRACT

Avian influenza such as H5N1 could infect humans and cause great health concern due to its high mortality rate. In March 2013, a novel reassortant avian-origin influenza A (H7N9) virus was found in several patients with severe respiratory illness in China. Since then, at least 82 people in China have been infected with this new virus and 17 have died from this virus. The question of how these people were infected with this virus and whether this virus will spread among people remains an urgent topic for research. This study took an early investigation of this virus by comparing the collected viral genome sequences of 2013 H7N9 in China against those of previous avian H7N9 and examined the receptor binding specificity of this new virus. This virus was found to be very different from the previous avian H7N9 viruses and surprisingly many of the internal proteins of 2013 H7N9 from the avian and human hosts in China were either identical or similar. Our analysis of the HA protein of this virus implied that the current strains of 2013 H7N9 in China displayed avian type receptors as their primary binding preference and human type receptors as secondary. For pandemic risk assessment, we also detected 23 mutations, including a few well known for host adaptation, in the HA1 domain of the HA protein from this virus. Each mutation was quantified for its impact on the receptor binding selection using a bioinformatics approach. Collectively these current mutations tended to decrease the HA binding affinity for avian type receptors and increase that for human type receptors, which could enhance the ability of this virus to infect humans.

Keywords: H7N9; Influenza; Receptor Specificity; Mutation

1. INTRODUCTION

The discovery of a novel reassortant avian-origin influenza A (H7N9) virus from several patients in China in March 2013 signaled a major global public health concern [1]. This new virus is continuing its spread in China, raising the total number of people affected to at least 82 including 17 deaths as indicated in a news report from <http://news.xinhuanet.com> on April 17, 2013. In general human influenza tends to cause mild symptoms, but avian influenza such as H5N1 is more deadly for people. Human infections with avian influenza are most likely after direct contact with infected poultry. Avian H7N9 virus did not infect humans before March 2013 and certain gene mutations need to occur in order for it to acquire the capability to infect humans and spread between people. Because of the fast evolution rate of influenza, it is critical for us to closely monitor any evidence that 2013 H7N9 is passing from person to person, which could dramatically increase its pandemic potential.

The influenza A virus genome is made of eight gene segments that encode 10 proteins: HA, NA, PB1, PB2, PA, NP, M1, M2, NS1, NS2. The two surface proteins HA and NA are responsible for the entry of the virus into the host cells and release of new generation of viral particles from host cells respectively. The remaining eight proteins are internal proteins that have different functions ranging from facilitating the reproduction of the virus to providing structural support to the viral envelop. The segmented structure of genome makes it easier to form novel influenza viruses from reassortment of different influenza viruses. The study in [1] suggested that this novel 2013 H7N9 virus in China is of avian origin. Its NA gene is similar to that of another H7N9 virus from Korea (A/wild bird/Korea/A14/2011), its HA gene is close to that of an H7N3 virus from Zhejiang Province in China (A/duck/Zhejiang/12/2011), and all its internal gene segments are very similar to those from avian H9N2 viruses, particularly a virus isolated from a bram-

bling in Beijing (A/brambling/Beijing/16/2012).

Although the host barrier for avian viruses to spread in humans needs further elucidation, the receptor binding specificity of HA is widely recognized as a major obstacle for direct avian to human transmission. In general, human influenza viruses tend to bind to SA α 2,6Gal receptors, whereas avian viruses favor SA α 2,3Gal receptors. Adaptation of avian virus to humans likely requires a switch in receptor binding specificity of the virus from avian-type to human-type.

Inspired by the work in [1], which used three 2013 H7N9 strains collected from three patients in China, this study employed the methodology developed in [2-17] to investigate this novel avian-origin H7N9 virus with five 2013 H7N9 strains in China. Our aim was to discover gene sequence difference between this new virus and the previous avian H7N9 and identify and then analyze mutations in the HA protein that could help this virus making its adaptation from avian to human hosts.

2. MATERIALS AND METHODS

2.1. Sequence Data

Protein sequences of influenza viruses used in this study were retrieved from the EpiFlu Database (<http://platform.gisaid.org>) of GISAID. All sequences were aligned with MAFFT [18] and phylogeny analysis of these sequences was conducted with MEGA [19]. The Meta information of the 2013 H7N9 sequences in China is listed in **Table 1**. A/Hangzhou/1/2013 does not have all the 10 proteins as others in the sequence collection of 2013 H7N9 strains in China. The study in [1] utilized the first three viral genome sequences in **Table 1** available at that time.

2.2. Informational Spectrum Method

The informational spectrum method (ISM) is a bioin-

formatics technique that can be used to analyze protein sequences [20]. The idea is to translate the protein sequences into numerical sequences based on electron-ion interaction potential (EIIP) of each amino acid. Then the Discrete Fourier Transform (DFT) can be applied to these numerical sequences, and the resulting DFT coefficients are used to produce the energy density spectrum. The informational spectrum (IS) comprises the frequencies and the amplitudes of this energy density spectrum. According to the ISM theory, the peak frequencies of IS of a protein sequence reflect its biological or biochemical functions. The ISM was successfully applied to quantify the effects of HA mutations on the receptor binding preference in [7,21,22].

3. RESULTS

3.1. Sequence Comparison

We compared five viral genome sequences of 2013 H7N9 in China with 13 sequences of previous avian H7N9 (**Figure 1**). It appeared that these five sequences

Table 1. Protein sequences of 2013 H7N9 in China.

ID	Protein
A/Shanghai/1/2013	HA, NA, PB1, PB2, PA, NP, M1, M2, NS1, and NS2
A/Shanghai/2/2013	HA, NA, PB1, PB2, PA, NP, M1, M2, NS1, and NS2
A/Anhui/1/2013	HA, NA, PB1, PB2, PA, NP, M1, M2, NS1, and NS2
A/Hangzhou/1/2013	HA
A/Chicken/Shanghai/S1053/2013	HA, NA, PB1, PB2, PA, NP, M1, M2, NS1, and NS2
A/Pigeon/Shanghai/S1069/2013	HA, NA, PB1, PB2, PA, NP, M1, M2, NS1, and NS2

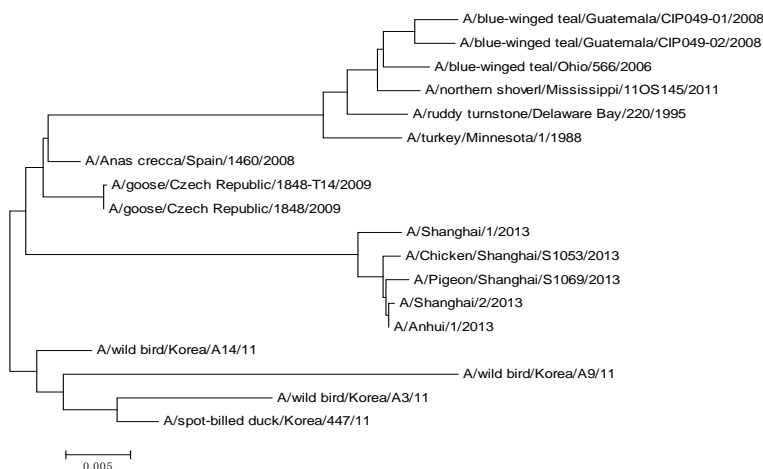


Figure 1. Polygenic tree of five viral genome sequences of 2013 H7N9 in China and those of previous avian H7N9.

were quite very different from the previous avian H7N9 influenza. It was interesting to note that A/Shanghai/1/2013, though a human strain, was not as similar to the other two human strains, A/Shanghai/2/2013 and A/Anhui/1/2013, as the two avian strains. In particular, A/Shanghai/1/2013 and A/Shanghai/2/2013 were actually collected from the same city Shanghai.

We also compared the individual protein sequences of five 2013 H7N9 strains in China with that of the consensus sequence of 13 previous avian H7N9 strains as

seen in **Figure 1 (Figure 2)**.

The polygenic trees in **Figure 2** implied that many of the internal proteins of avian and human 2013 H7N9 in China were either identical or similar, but remained very different from those of previous avian H7N9. The most diverse protein was the surface protein HA. There was a deletion in the stalk region of NA proteins [2] and a deletion at the C terminal of NS1 proteins of avian and human 2013 H7N9 in China as seen from the sequence display below (**Table 2**).

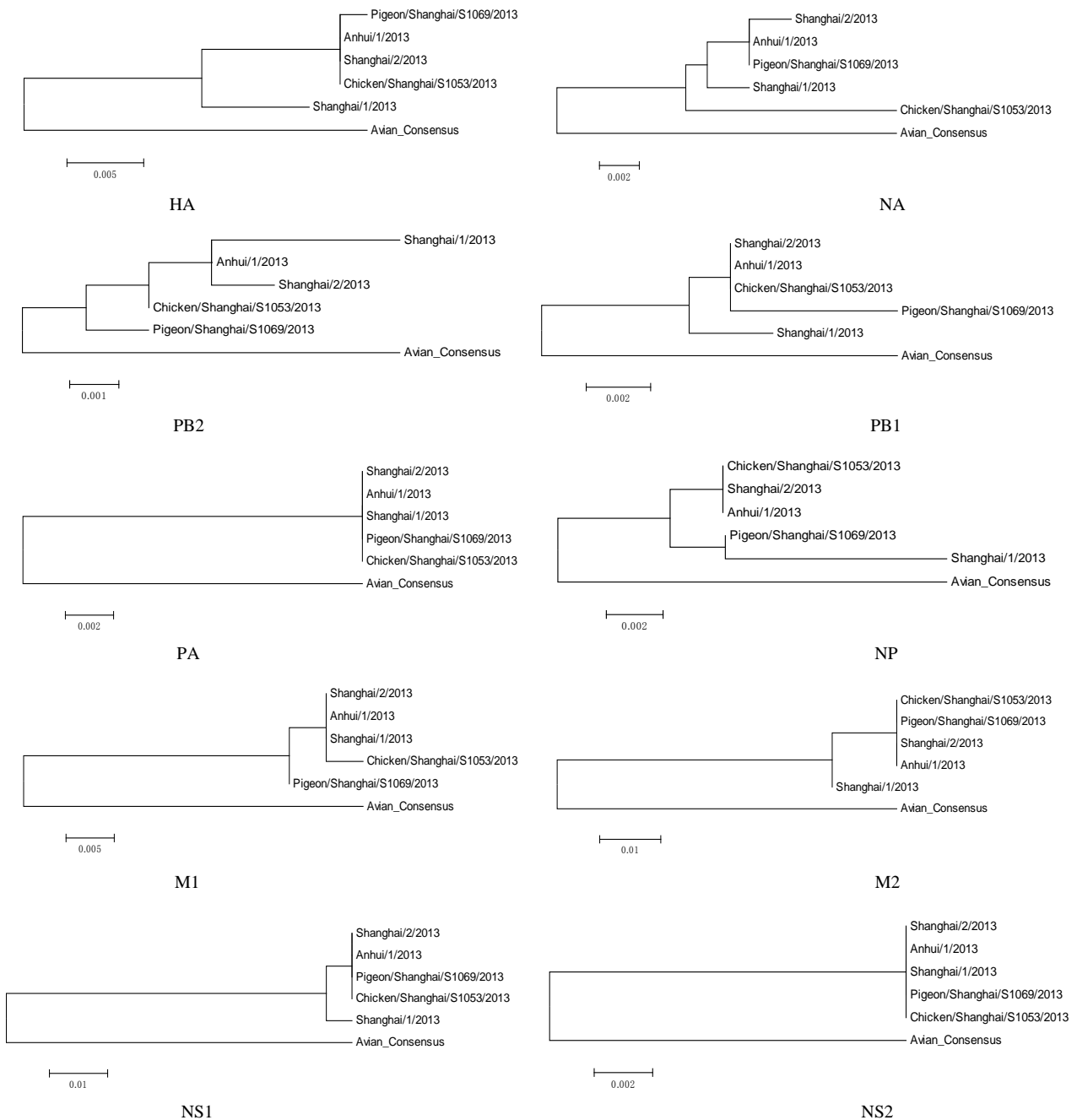


Figure 2. Polygenic trees of proteins of five 2013 H7N9 strains in China and those of the consensus of previous avian H7N9.

3.2. Receptor Affinity

The HA protein of influenza is essential for the viral entry into the host cells and its receptor binding specificity is an important property of its function. In this part of study, we also included an extra HA protein sequence of 2013 H7N9 in China: A/Hangzhou/1/2013.

3.2.1. Finding Mutations in the HA1 Domain of the HA Protein of 2013 H7N9 in China

To render the whole picture and comparison of the sequences of HA1 of 2013 H7N9 in China with a consensus sequence made of previous avian H7N9, the alignment of them is presented in **Table 3**. Based on the sequence alignment, we were able to identify mutations in the HA1 domain of the HA protein of 2013 H7N9 in China (**Tables 4** and **5**). Two critical mutations T160A and Q226L were discovered in [1].

3.2.2. Measuring the Effects of Mutations in HA on the Receptor Selection of 2013 H7N9 in China

The ISM technique was applied to the mutations discovered in Section 3.2.1. We used 51 HA protein sequences of human H7 HA collected prior to 2013 to calculate the primary CIS frequency of F(0.328) and 13 HA protein sequences of avian H7N9 collected prior to 2013 to calculate the primary CIS frequency of F(0.285). Therefore frequency F(0.328) was for binding human receptors whereas frequency F(0.285) was for avian receptors. The primary and secondary CIS frequencies were identified for the four human HA proteins of 2013 H7N9 (Shanghai/1/2013, Shanghai/2/2013, Anhui/1/2013, and Hangzhou/1/2013) and the two avian HA proteins of 2013 H7N9 (Chicken/Shanghai/S1053/2013 and Pigeon/hanghai/S1069/2013), both of which were F(0.285) and F(0.328). A study in [11] indicated that many influenza viruses tend to have dual receptor binding preference demonstrated by their primary and secondary CIS fre-

quencies. Furthermore the IS of the HA protein of Hangzhou/1/2013 and that with a mutation I266Q were computed. To offer comparison with other H7 HA proteins, the IS of the HA protein of A/mallard/Netherlands/9/2005/H7N7 was also provided (**Figure 3**).

The net effect of mutation Q226I on the HA protein of Hangzhou/1/2013 was to change the IS amplitude at F(0.285) from 6.7662 to 6.9940 and that at F(0.326) from 6.4342 to 6.4703. The HA1 sequences of Anhui/1/2013 and Hangzhou/1/2013 differed only at position 226, where the first has 226I and the second had 226L. Because EIIP value for both I and L is the same, mutation Q226L on Anhui/1/2013 had the same effect as mutation Q226I on Hangzhou/1/2013. Mutation Q226L is also known to be associated with transmission of avian H5N1 viruses by respiratory droplets in ferrets [23,24].

Besides Q226L, T160A was another mutation discovered in the HA protein of 2013 H7N9 in China [1]. A report from [25] suggested that double mutations S158N and T160A in the HA protein of H5N1 resulted in increased binding to human type receptors without a loss of binding to avian type receptors. Here we applied mutations S158N and T160A to Hangzhou/1/2013, and found that the IS amplitude at F(0.285) decreased by 0.0411 (0.6%) and that at F(0.326) increased by 0.4136 (6.4%). Our calculation thus showed that these two mutations had the same HA binding shift effect on Hangzhou/1/2013 as on the H5N1 virus.

Our next task was to calculate the IS amplitude change caused by each individual mutation found in Section 3.2.1 (**Table 6**). We selected the HA protein of Hangzhou/1/2013 as one representative of the six HA proteins of the avian and human 2013 H7N9 in China and applied these mutations to this HA protein. Collectively they tended to enhance the receptor affinity for human type receptors while reduce the affinity for avian type receptors since the average IS amplitude change was -0.0988 (1.41%) at F(0.295) and 0.0293 (0.045%) at F(0.326).

Table 2. Deletion of NA and NS1 proteins from 2013 H7N9 in China when compared to those from previous avian H7N9.

The NA protein sequences from positions 1-120:

```
>Avian_Consensus
MNPQKILCTSATAIVIGTIAVLIGIANLGLNIGLHLKPCNSCSHSQPEATNASQTIINNNYNETNITQISNTNIQMEERASREFNNTKGLCTINSWHIYGKDNAVRIGENSDVLVTRE
> Chicken/Shanghai/S1053/2013
MNPQKILCTSATAIIGAIIVLIGIANLGLNIGLHLRPSNCNSHSQPETTNTSQTIVINNNYNETNI-----TNIQMEERTSRNFNNTKGLCTINSWHIYGKDNAVRI GESSDVLVTRE
1
```

The whole NS1 protein sequences:

```
>Avian_Consensus
MDSNTVSSFOVDCFLWHVRRKRFADQELGDPFLDRLRRDQKSLRGRGSLGLDITETATRAGKQIVERILEEESDEALKMTIASVPASRYLTDMTLEMSRDWFLMMPKQKVAGSLCIR
MDQAIMDKNILKANFSVIFDRLETLILLRAFTEEGAIVGEISPLPSLPGHTDEDVKNAIEVLIGGFENWNTVVRVSETLQRFARWSSNEDGRPPLPPKQKRKMARTIESEV
>Chicken/Shanghai/S1053/2013
MDSNTVSSFOVDCFLWHVRRKRFADQEMGDPFLDRLRRDQKSLRGRSSTLGLDITRTATREGKHIIVERILEEESDEAFKMSIASVPAPRYLTDMTLEMSRDWMLIPKQKITGSLCIR
MDQAIIVDKNITLKANFSVIFNRLEALILLRAFTEEGAIVGEISPLPSLPGHTDKDKNAIEILIGGFENWNTVVRVSETLQRFARWSSDEDEGRSPLSTK-----
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Table 3. Alignment of HA1 sequences of six 2013 H7N9 strains in China and the consensus HA1 sequence of previous avian H7N9.

Avian_Consensus	DKICLGHHAVSNGTKVNTLTERGVEVVNATETVERTNIPRICSKGKRTVDLGQCGLLGTI
Chicken/Shangha	DKICLGHHAVSNGTKVNTLTERGVEVVNATETVERTNIPRICSKGKRTVDLGQCGLLGTI
Pigeon/Shanghai	DKICLGHHAVSNGTKVNTLTERGVEVVNATETVERTNIPRICSKGKRTVDLGQCGLLGTI
Shanghai/1/2013	DKICLGHHAVSNGTKVNTLTERGVEVVNATETVERTNIPRICSKGKRTVDLGQCGLLGTI
Shanghai/2/2013	DKICLGHHAVSNGTKVNTLTERGVEVVNATETVERTNIPRICSKGKRTVDLGQCGLLGTI
Anhui/1/2013	DKICLGHHAVSNGTKVNTLTERGVEVVNATETVERTNIPRICSKGKRTVDLGQCGLLGTI
Hangzhou/1/2013	DKICLGHHAVSNGTKVNTLTERGVEVVNATETVERTNIPRICSKGKRTVDLGQCGLLGTI *****:*****
Avian_Consensus	TGPPQCDQFLEFSADLI IERREGSDVCYPGKFVNEEALRQILRESGGIDKETMGFTYSGI
Chicken/Shangha	TGPPQCDQFLEFSADLI IERREGSDVCYPGKFVNEEALRQILRESGGIDKEAMGFTYSGI
Pigeon/Shanghai	TGPPQCDQFLEFSADLI IERREGSDVCYPGKFVNEEALRQILRESGGIDKEAMGFTYSGI
Shanghai/1/2013	TGPPQCDQFLEFSADLI IERREGSDVCYPGKFVNEEALRQILRESGGIDKEAMGFTYSGI
Shanghai/2/2013	TGPPQCDQFLEFSADLI IERREGSDVCYPGKFVNEEALRQILRESGGIDKEAMGFTYSGI
Anhui/1/2013	TGPPQCDQFLEFSADLI IERREGSDVCYPGKFVNEEALRQILRESGGIDKEAMGFTYSGI
Hangzhou/1/2013	TGPPQCDQFLEFSADLI IERREGSDVCYPGKFVNEEALRQILRESGGIDKEAMGFTYSGI *****:*****
Avian_Consensus	RTNGATSACRRSGSSFYAEMKWLLSNTDAAFPQMTKSYKNRNDPALIIWGIHHSGSTT
Chicken/Shangha	RTNGATSACRRSGSSFYAEMKWLLSNTDAAFPQMTKSYKNTRKSPALIVWGIHHSVSTA
Pigeon/Shanghai	RTNGATSACRRSGSSFYAEMKWLLSNTDAAFPQMTKSYKNTRKSPALIVWGIHHSVSTA
Shanghai/1/2013	RTNGATSSCRRSGSSFYAEMKWLLSNTDAAFPQMTKSYKNTRKSPALIVWGIHHSGSTA
Shanghai/2/2013	RTNGATSACRRSGSSFYAEMKWLLSNTDAAFPQMTKSYKNTRKSPALIVWGIHHSVSTA
Anhui/1/2013	RTNGATSACRRSGSSFYAEMKWLLSNTDAAFPQMTKSYKNTRKSPALIVWGIHHSVSTA
Hangzhou/1/2013	RTNGATSACRRSGSSFYAEMKWLLSNTDAAFPQMTKSYKNTRKSPALIVWGIHHSVSTA *****:*****:*****:***** **:
Avian_Consensus	EQTKLYGSGSKLITVGSSNYQQSFVSPGARPQVNGQSGRIDFHWWLLNPNNDTVTFNFNG
Chicken/Shangha	EQTKLYGSGNKLVTVGSSNYQQSFVSPGARPQVNGLSGRIDFHWWMLNPNNDTVTFNFNG
Pigeon/Shanghai	EQTKLYGSGNKLVTVGSSNYQQSFVSPGARPQVNGLSGRIDFHWWMLNPNNDITTFNFNG
Shanghai/1/2013	EQTKLYGSGNKLVTVGSSNYQQSFVSPGARTQVNGQSGRIDFHWWMLNPNNDTVTFNFNG
Shanghai/2/2013	EQTKLYGSGNKLVTVGSSNYQQSFVSPGARPQVNGLSGRIDFHWWMLNPNNDTVTFNFNG
Anhui/1/2013	EQTKLYGSGNKLVTVGSSNYQQSFVSPGARPQVNGLSGRIDFHWWMLNPNNDTVTFNFNG
Hangzhou/1/2013	EQTKLYGSGNKLVTVGSSNYQQSFVSPGARPQVNGISGRIDFHWWMLNPNNDTVTFNFNG *****:*****:*****:*****:*****:*****
Avian_Consensus	AFIAPDRASFLRGKSMGIQSGVQVDASCEGDCYHSGGTIIISNLPFQINIDRAVVKCPRVY
Chicken/Shangha	AFIAPDRASFLRGKSMGIQSGVQVDANCEGDCYHSGGTIIISNLPFQINIDRAVVKCPRVY
Pigeon/Shanghai	AFIAPDRASFLRGKSMGIQSGVQVDANCEGDCYHSGGTIIISNLPFQINIDRAVVKCPRVY
Shanghai/1/2013	AFIAPDRASFLRGKSMGIQSGVQVDADCEGDCYHSGGTIIISNLPFQINIDRAVVKCPRVY
Shanghai/2/2013	AFIAPDRASFLRGKSMGIQSGVQVDANCEGDCYHSGGTIIISNLPFQINIDRAVVKCPRVY
Anhui/1/2013	AFIAPDRASFLRGKSMGIQSGVQVDANCEGDCYHSGGTIIISNLPFQINIDRAVVKCPRVY
Hangzhou/1/2013	AFIAPDRASFLRGKSMGIQSGVQVDANCEGDCYHSGGTIIISNLPFQINIDRAVVKCPRVY *****:*****:*****:*****:*****
Avian_Consensus	KQESLLLATGMKNVPEIPKGR
Chicken/Shangha	KQRSLLLATGMKNVPEIPKGR
Pigeon/Shanghai	KQRSLLLATGMKNVPEIPKGR
Shanghai/1/2013	KQRSLLLATGMKNVPEIPKGR
Shanghai/2/2013	KQRSLLLATGMKNVPEIPKGR
Anhui/1/2013	KQRSLLLATGMKNVPEIPKGR
Hangzhou/1/2013	KQRSLLLATGMKNVPEIPKGR **:***** **

Table 4. Amino acids at critical sites (H3 numbering) for receptor selection in the HA protein. The distances in the table represent the Hamming distances between the consensus of HA sequences of previous avian H7N9 and others based on the amino acids at the 17 sites in the table.

Position	98	136	153	183	186	190	193	194	195
H1N1 human consensus	Y	S	W	H	P	D	A	L	Y
H1N1 California/04/2009	Y	T	W	H	S	D	S	L	Y
H5N1 human consensus	Y	T	W	H	N	E	K	L	Y
H7N9 avian consensus	Y	T	W	H	G	E	K	L	Y
Chicken/Shanghai/S1053/2013	Y	T	W	H	V	E	K	L	Y
Pigeon/Shanghai/S1069/2013	Y	T	W	H	V	E	K	L	Y
Shanghai/1/2013	Y	T	W	H	G	E	K	L	Y
Shanghai/2/2013	Y	T	W	H	V	E	K	L	Y
Anhui/1/2013	Y	T	W	H	V	E	K	L	Y
Hangzhou/1/2013	Y	T	W	H	V	E	K	L	Y
Position	196	216	221	222	225	226	227	228	Dist
H1N1 human consensus	H	E	P	K	D	Q	E	G	9
H1N1 California/04/2009	Q	E	P	K	D	Q	E	G	8
H5N1 human consensus	Q	K	S	K	G	Q	S	G	4
H7N9 avian consensus	G	S	P	Q	G	Q	S	G	0
Chicken/Shanghai/S1053/2013	G	S	P	Q	G	L	S	G	2
Pigeon/Shanghai/S1069/2013	G	S	P	Q	G	L	S	G	2
Shanghai/1/2013	G	S	T	Q	G	Q	S	G	1
Shanghai/2/2013	G	S	P	Q	G	L	S	G	2
Anhui/1/2013	G	S	P	Q	G	L	S	G	2
Hangzhou/1/2013	G	S	P	Q	G	I	S	G	2

Table 5. Additional mutations found in the HA proteins of 2013 H7N9 in China.

Position	47	122	138	171	173	174	179	186	189	199
H7N9 avian consensus	V	T	A	P	N	D	I	G	T	S
Chicken/Shanghai/S1053/2013	I	A	A	T	K	S	V	V	A	N
Pigeon/Shanghai/S1069/2013	I	A	A	T	K	S	V	V	A	N
Shanghai/1/2013	I	A	S	T	K	N	V	G	A	N
Shanghai/2/2013	I	A	A	T	K	S	V	V	A	N
Anhui/1/2013	I	A	A	T	K	S	V	V	A	N
Hangzhou/1/2013	I	A	A	T	K	S	V	V	A	N
Position	202	221	236	243	277	283	298	312	315	326
H7N9 avian consensus	I	P	L	V	S	H	N	E	M	N
Chicken/Shanghai/S1053/2013	V	P	M	V	N	H	D	R	L	I
Pigeon/Shanghai/S1069/2013	V	P	M	I	N	H	D	R	L	I
Shanghai/1/2013	V	T	M	V	D	Y	D	R	L	I
Shanghai/2/2013	V	P	M	V	N	H	D	R	L	I
Anhui/1/2013	V	P	M	V	N	H	D	R	L	I
Hangzhou/1/2013	V	P	M	V	N	H	D	R	L	I

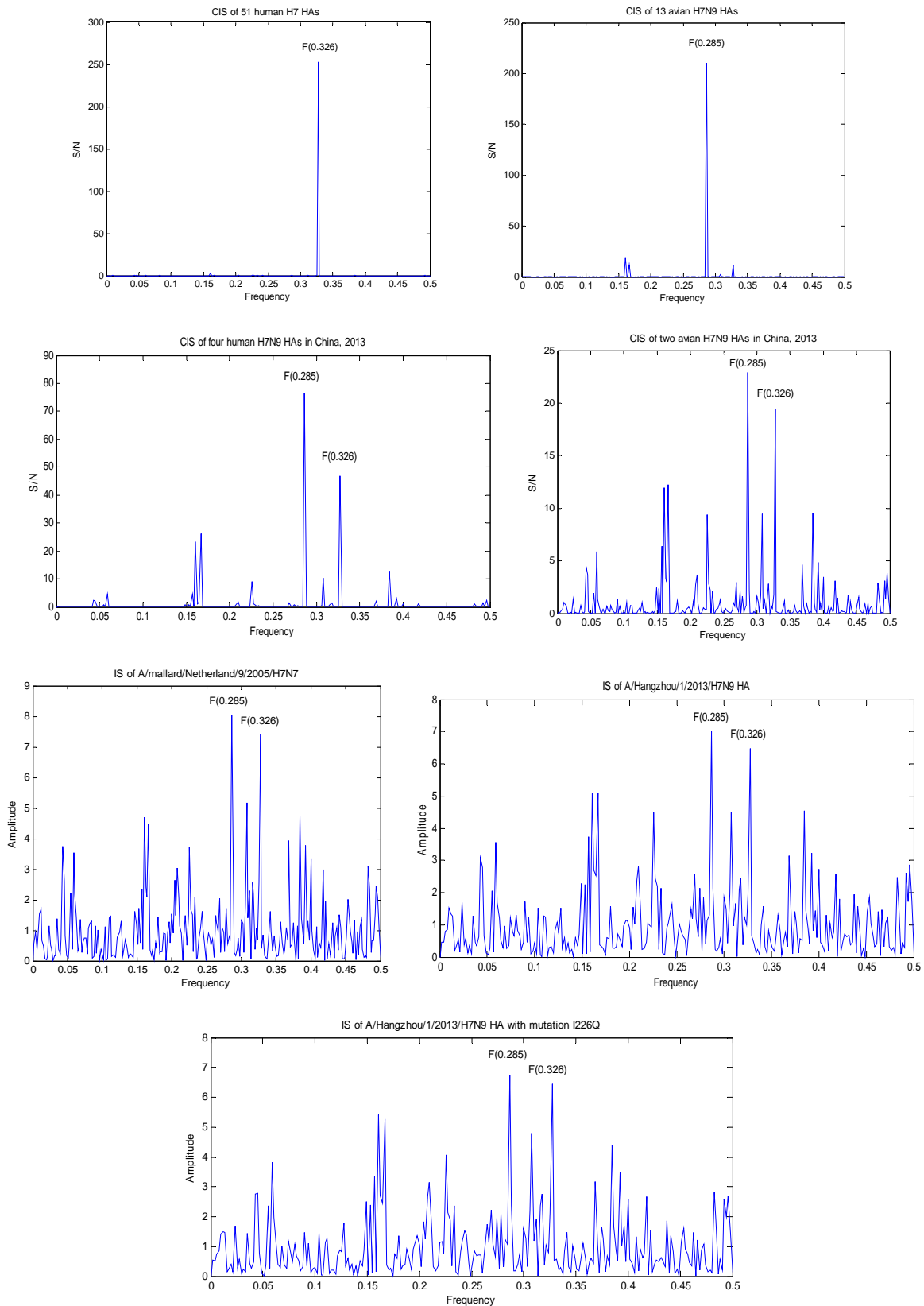


Figure 3. CIS of 51 human H7 HA proteins, CIS of 13 avian H7N9 HA proteins, CIS of four HA proteins of human 2013 H7N9 in China, CIS of two HA proteins of avian 2013 H7N9 in China, IS of the HA protein of A/Hangzhou/1/2013 and IS of that with mutation I226Q, and IS of the HA protein of A/mallard/Netherlands/9/2005/H7N7.

Table 6. Calculation of the IS amplitude change of HA of A/Hangzhou/1/2013 caused by the mutations at the primary and second binding frequencies.

Mutation	IS amplitude change at F(0.285)	IS amplitude change at F(0.326)
V47I	0.0219	-0.0357
T122A	0.0667	-0.3675
A138S	0.1081	-0.011
P171T	0.5041	-0.2764
N173K	-0.2044	0.2477
D174S	-0.1129	0.1636
D174N	0.3165	-0.2028
I179V	0.0003	0.043
G186V	1E-04	0.0018
T189A	-0.266	-0.1016
S199N	-0.5828	0.5208
I202V	0.0266	-0.0318
P221T	0.0103	0.35
Q226I	0.2278	0.0361
L236M	-0.5944	-0.5721
V243I	-0.0352	0.0211
S277N	-0.7631	0.7874
S277D	-0.5364	0.4789
H283Y	-0.1853	-0.1261
N298D	0.1449	-0.0874
E312R	0.1755	-0.2925
M315L	-0.5832	0.1453
N326I	-0.0135	-0.0223
Average	-0.0988	0.0293

To visualize the impact of these mutations, we plotted the numerical values of IS amplitude change in **Figure 4**. Mutations S199N, S277N, and S277D clearly favored human receptor binding and reduced avian receptor binding and all of these three mutations mutated the amino acid S. The famous mutation Q226I also found in [1] only generated moderate change that increased both human and avian receptor binding. Another interesting mutation was L236M that reduced both of avian and human binding greatly.

4. CONCLUSION

The present study made use of the current available viral genome sequences of 2013 H7N9 in China. Our goal was to make an early investigation of this novel virus. Se-

quence comparison revealed that this new virus was quite different from previous avian H7N9, but many of the internal proteins of 2013 H7N9 in China were either identical or similar. For pandemic risk assessment, the HA protein of this new virus was examined due to its critical role in the adaptation to human hosts. Our analysis implied that the current strains of 2013 H7N9 in China exhibited avian type receptors as their primary binding preference and human type receptors as secondary. We found 23 mutations in the HA1 domain of the HA protein of this virus. Each of these mutations was measured for its effect on receptor selection. Three mutations S199N, S277N, and S277D in the HA protein of 2013 H7N9 in China showed evident power to increase the binding affinity for human type receptors and at the

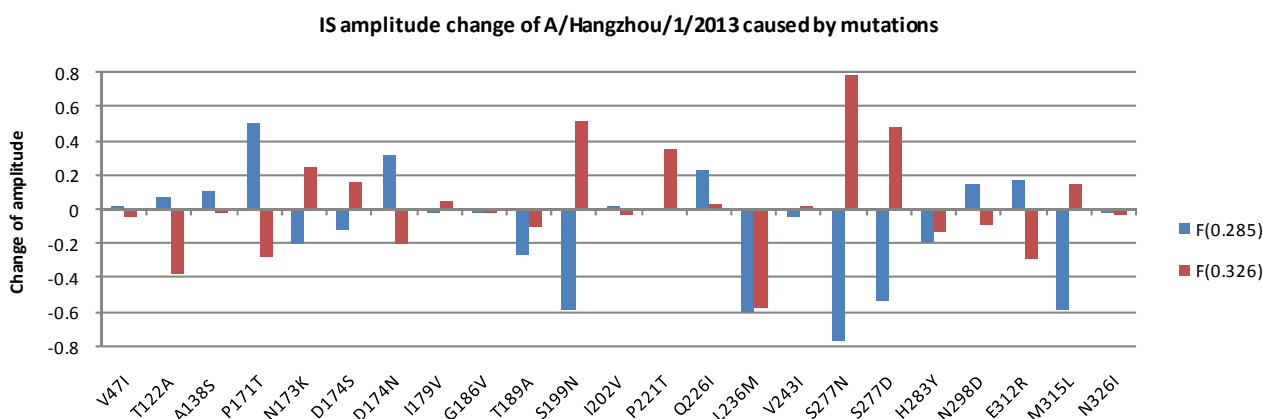


Figure 4. IS amplitude change of A/Hangzhou/1/2013 caused by 23 mutations in the HA proteins of 2013 H7N9 in China.

same time to reduce the affinity for avian type receptors. Collectively these mutations enhanced the binding to human type receptors whereas decreased the binding to avian type. Our findings suggested the current mutation pattern of this new virus contributed to its continued adaptation to human hosts.

5. ACKNOWLEDGEMENTS

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